Optimized production, quality control and biological assessment of ⁶⁸Ga-bleomycin as a possible PET imaging agent

H.R. Tayeri¹, E. Sattarzadeh Khameneh², S. Zolghadri², S. Kakaei^{3*}, D. Sardari¹

¹Department of Nuclear Engineering, Science and Research Branch, Islamic Azad University, Tehran, Iran ²Nuclear Fuel Cycle Research School, Nuclear Science and Technology Research Institute (NSTRI), , Tehran, Iran ³Radiation Application Research School, Nuclear Science and Technology Research Institute, Tehran, Iran

ABSTRACT

Background: Recently, several diagnostic radiopharmaceuticals including bleomycin (BLM) derivatives have been developed. Due to the suitable physical and chemical characteristics of ⁶⁸Ga as a radionuclide for PET imaging, in this study, optimized production, quality control and preclinical evaluation of ⁶⁸Ga-BLM as a new PET imaging agent is reported for the first time. Materials and Methods: Labeling of BLM with ⁶⁸Ga was performed using ⁶⁸Ge/⁶⁸Ga generator. Experiments were carried out by changing BLM concentration, temperature and pH of the reaction to determine the optimum parameters while the radiochemical purity was checked by radio thin layer chromatography at different times post labeling. Stability of the radiolabeled complex was studied at room temperature and in human serum at 37 °C. Biodistribution of the complex in BALB/c mice was assessed after intravenous injection and by counting the activity of each organ. Also, images were acquired up to 120 min by dual-head SPECT system. Results: The purity of this complex >96% (ITLC). At the optimized conditions for preparation of ⁶⁸Ga-BLM (pH= 3.5-4, temperature = 90 ^oC, reaction time = 15 minutes and ligand concentration of 1 mg/ml), the special activity of the labeled BLM reached to around 17.5 GBq/mmol. Biodistribution study showed significant accumulation of radioactivity in lung and bladder that was different pharmacokinetic compared to free ⁶⁸Ga cation. *Conclusion:* Results show that ⁶⁸Ga-BLM can be prepared in high radiochemical purity and high special activity only in 15 minutes and totally can be considered as a high potential agent for PET imaging.

Keywords: Bleomycin, Gallium-68, radiolabeled compound, PET.

Original article

*Corresponding authors:

Saeed Kakaei, Ph.D.

E-mail: Skakaei@aeoi.org.ir

Revised: July 2019 Accepted: August 2019

Int. J. Radiat. Res., April 2020;

18(2): 235-241

DOI: 10.18869/acadpub.ijrr.18.2.235

INTRODUCTION

Early detection of cancers can greatly increase the chance of successful treatment and radiolabeled tracers can play an important role for primitive diagnosis of tumors. Positron emission tomography (PET) has rapidly become an indispensable tool for clinicians in the locating and staging of tumors, as well as

measuring their response to interventional therapies. PET experienced rapid development and has become an accepted approach for clinical routine diagnostics. Therefore, the expansion of new radiopharmaceuticals for development of clinical PET is very essential ⁽¹⁾.

Excellent physical specifications of 68 Ga such as relatively short half-life of 67.71 min, positron emission of 89 % and low abundance of 1077

Tayeri et al. / 68 Ga-bleomycin: New agent PET imaging

keV photon emission (3.22 %) as well as it's availability in the form of 68Ge/68Ga generator at a reasonable cost make this radioisotope an developing attractive choice for radiopharmaceuticals specially for the countries with limited or no cyclotron facilities (2,3). Gallium-68 has high tumor affinity and it is being used in nuclear medicine for detection and imaging of malignant and inflammatory sites in the body for a short time. The number of publications related to basic and clinical researches of 68Ga-radiopharmaceutical has drastically promoted during recent years (1,4-7).

Bleomycins (BLMs) are family glycopeptide antibiotics that have potent antitumor activity against a range of lymphomas, head and neck cancers and germ-cell tumors and therefore, are widely used for chemotherapy of cancers. The mechanisms of BLM induced cytotoxicity have been studied (8,9) and when in presence of Fe⁺² ion and oxygen, it transforms into its active form and binds to DNA molecules. Several structure variations of BLMs have been identified from fermentation broths, primarily differing at the C-terminus of the glycopeptide. Structurally and biosynthetically related to the BLMs are the phleomycins (PLMs), such as PLM 12 or PLM D1 (10,11), and tallysomycins (TLMs), such as TLM S2B and TLM S10B (12,13) (figure 1).

During the past three decades, complexes of BLM with radioactive elements as a

radiopharmaceutical and their body distribution and resultant biological effects, were intensively studied. Several radiolabeled BLM derivatives contain indium-111 ⁽¹⁴⁾, cobalt-57 ⁽¹⁵⁾, technetium-99m ⁽¹⁶⁾, radioferric salts ⁽¹⁷⁾, rhodium-105 ⁽¹³⁾ and radiolanthanides such as lutetium-177, samarium-153 and holmium-166 ⁽¹⁶⁻¹⁸⁾ have been developed for imaging and/or therapy of neoplastic tissues.

However, the positron-emitting gallium (III) radioisotopes, 66Ga3+ and 68Ga3+, have been proposed for applications in PET imaging, according to the best of our knowledge, preparation, quality control and biological of ⁶⁸Ga-BLM complex have not been reported until now. Therefore, in this research study, due to the suitable physical characteristics of 68Ga and desirable tumor seeking properties of BLM, preparation optimized conditions for ⁶⁸Ga-BLM complex were investigated. Different affecting parameters on the labeling yield such as pH, temperature, and time and ligand concentration were studied. The quality control of the complex was done by radio thin layer chromatography (RTLC) method and the stability and biodistribution of the radiolabeled compound were checked in normal animal model. Also, the imaging of the animal was acquired by a dual-head SPECT system after injection of the radiolabeled compound to the animals.

Figure 1. Structures of bleomycinic acid, bleomycin A2, bleomycin B2, phleomycin 12, phleomycin D1, and tallysomycin S2B, and tallysomycin S10B.

MATERIALS AND METHODS

⁶⁸Ge/⁶⁸Ga generator with the nominal activity of 50 mCi was provided from Pars Isotope Co. (Tehran, Iran) and its chemical purity was check by an ICP-OES spectrometer (Varian Co., model Turbo-AX-150-Liberty). BLM sulfate purchased from Merck (Darmstadt, Germany). buffer acetate. ammonium methanol and other chemical reagents were obtained from Sigma Aldrich (UK). A high purity germanium (HPGe) detector coupled with a Canberra™ (model GC1020-7500SL) multichannel analyzer used for was radionuclidic purity assessment. Also, radiochemical purity analysis was done by a bioscan AR-2000 radio TLC scanner instrument (Bioscan, Europe Ltd CO., France) Whatman No. 2 paper (Whatman, Maidstone, UK). Student T-test was used to compare the obtained data and the final values were expressed as the mean ± standard deviation (Mean ± SD). Statistical significance was defined as P<0.05.

Preparation and quality control of 68GaCl₃

The QC/QA protocols for used in this study have been reported elsewhere (21,4). To prepare ⁶⁸GaCl₃ solution, ⁶⁸Ge/⁶⁸Ga generator was eluted by 2 mL HCl (0.6 M) and gathered in separate fractions. The first 0.5 mL of the eluate was discarded and the next 1.5 ml was used for labeling purposes after the activity measurement. The radionuclidic purity of the product was investigated by spectrometry for 1000 s. The sample was for counted 48 h later breakthrough of ⁶⁸Ge measurement in the sample. Radiochemical purity of 68GaCl₃ solution was checked in 10 % ammonium acetate/methanol mixture on silica gel sheets and in 10 mM DTPA solution (pH = 4) on Whatman No. 2 paper.

Radiolabeling of BLM with 68GaCl₃

BLM powder was dissolved in pure water to prepare the stock solution with concentration of 1 mg/mL. ⁶⁸GaCl₃ solution was added to the 10-mL conical vial and evaporated by slight heating under nitrogen gas flow. Different *Int. J. Radiat. Res., Vol. 18 No. 2, April 2020*

volumes of BLM solution (0.5–2 mmol) were added to the vials. To determine the optimum parameters for reaction, temperature and pH of the reaction changed from 25–100 °C and 1–5, respectively. While, the maximum reaction time considered being 1 h, the radiochemical purity of the complex was checked at different times post labeling. Radiochemical purity tests were carried out by RTLC with various mobile phase mixtures including 10% Ammoniumacetate:Methanol (1:1); H₂O:Methanol: Acetic acid (4:4:2).

Stability tests of 68Ga-BLM

The stability of 68 Ga-BLM was performed at room temperature up to 120 min post preparation by checking the radiochemical purity of the complex at the specified intervals. Also, the stability of the complex in human serum was done by adding a 37 MBq of 68 Ga-BLM to 500 μ L of freshly prepared human serum following by incubation of the mixture at 37 °C for 48 h. Finally, aliquots were analyzed by ITLC method at various intervals after preparation.

Biological evaluation of 68 Ga-BLM in BALB/c mice

The protocol of the study has been approved by the institutional review board (Nuclear Science and Technology Research Institute (NSTRI)- License No.18, date 2013). The female BALB/c mice of age 28±4 days weighting 18.5-22.7 g was prepared from Pasteur institute (Tehran, Iran) and kept at routine day/night light program under common rodent diet pellets. 100 μ L of ⁶⁸GaCl₃ and ⁶⁸Ga-BLM (~ 37 MBq) solutions was intravenously injected into the mice through their tail veins. The mice sacrificed at the selected intervals (15, 30, 45 and 60 min) after injection (n=4). The tissues including liver, lung, spleen, muscle, skin, heart, kidney, colon, colon content, and bladder were rinsed with normal saline and weighed and their activities were determined using the HPGe detector. Finally, the activity concentration of each organ (A) was computed according to equation (1) (22).

$$A = \frac{N}{\epsilon \gamma \, t_s \, m \, k_1 \, k_2 \, k_3 \, k_4 \, k_5} \tag{1}$$

where N is the corrected net peak area of the

corresponding photopeak; ϵ is the efficiency at photopeak energy, γ is the emission probability of the gamma line corresponding to the peak energy, ts is the live time of the sample spectrum collection in seconds, m is the mass (kg) of the measured sample, k1 , k2, k3, k4 and k5 are the correction factors for the nuclide decay from the time the sample was collected to the start of the measurement, the nuclide decay during counting period, self-attenuation in the measured sample, pulses loss due to random summing and the coincidence, respectively.

Imaging studies

 $100~\mu L$ of 68 Ga-BLM solution (37 MBq) was injected intravenously to normal mice through their tail veins for imaging studies. PET images were taken at 30 and 120 min post administration by a dual-head SPECT system. The distances of mice to high-energy septa were 12 cm. The useful field of view (UFOV) was 540 mm×400 mm.

Statistical analysis

All values were expressed as mean \pm standard deviation and the data were compared using Student's T-test. Once a t value was calculated, the p-value was determined using the table of values from student's t distribution. Statistical significance was defined as P < 0.05.

RESULTS

Preparation and quality control of 68GaCl₃

The radionuclidic purity assessment 68GaCl₃ solution was done bv gamma spectrometry method demonstrated the presence of 511 and 1077 keV, all originating from ⁶⁸Ga. Also, the radiochemical purity of the sample used in this study was checked by ITLC method resulted in the radiochemical purity of more than 99.8%.

Radiolabeling of BLM with 68GaCl₃

To achieve optimized conditions for radiolabeling of BLM with ⁶⁸Ga, various experiments were done and the parameters including pH of reaction (1-5), temperature (25, 50, 75, 90 and 100°C), ligand concentration (0.5,

1, 1.5 and 2 mg), and reaction time (up to 1 h) were changed. While, 10% ammonium acetate: methanol (1:1) mixture was considered as a better mobile phase for control of the radiochemical purity, the optimization process was checked by this mobile phase. RTLC tests of the produced complex showed two distinct radio peaks at $R_{\rm f}$ s of 0.10 and 0.80 as indicated in figure 2.

Increasing the temperature of complex mixture during reaction time to 90°C increased the yield, which remained constant for temperatures up to 90-100 °C (figure 3). The best pH for labeling was found 3.5-4 at the temperature of 90 °C as indicated in figure 4. At the optimum reaction temperature and pH, the yield reached a maximum within 15 minutes which remained approximately constant for longer reaction times (figure 5).

Finally, the optimized conditions for radiolabeling of higher than 96 % were pH of 3.5 -4, temperature of 90 °C and the reaction time of 15 minutes that results in the special activity of around 17.5 GBq/mmol for ⁶⁸Ga-BLM.

Stability tests of 68Ga-BLM

The stability tests of the radiolabeled complex including stability at room temperature and stability in human serum at 37 °C were studied by ITLC method. The results showed that the radiochemical purity of ⁶⁸Ga-BLM was greater than 96 % after 120 min of preparation both at room temperature and in human serum at 37 °C.

Biological evaluation of ⁶⁸GaCl₃ and ⁶⁸Ga-BLM in BALB/c mice

Biodistribution of 68 GaCl $_{3}$ and 68 Ga-BLM was studied in BALB/c mice at 15, 30, 45 and 60 min post injection (figures 6 and 7).

Imaging studies

PET images were taken from normal BALB/c mice at 30 and 120 min post injection of ⁶⁸Ga-BLM solution. As shown in Fig. 8, the radiolabeled complex is mainly washed out from the urinary tract and the most accumulation occurred in bladder and the major remained activity is accumulated in lung and excretory organs.

Int. J. Radiat. Res., Vol. 18 No. 2, April 2020

Tayeri et al. / 68 Ga-bleomycin: New agent PET imaging

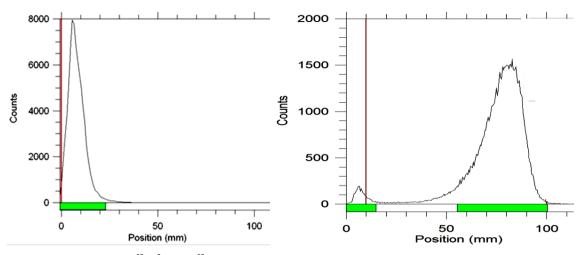


Figure 2. RTLC chromatogram of ⁶⁸Ga³⁺ (left), ⁶⁸Ga-BLM (right) in 10 % ammonium acetate/methanol mixture on silica gel sheets at optimized condition.

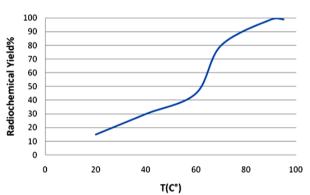
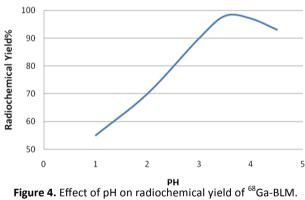


Figure 3. Effect of temperature on radiochemical yield of ⁶⁸Ga-BLM.



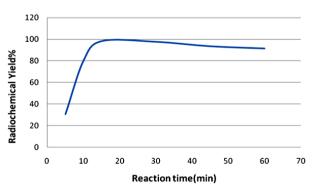
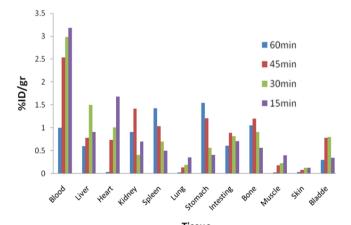


Figure 5. Effect of reaction time on radiochemical yield of ⁶⁸Ga-BLM.



Tissue **Figure 6.** Biodistribution of ⁶⁸GaCl3 in Balb/c mice up to 60 min after injection.

Taveri et al. / 68 Ga-bleomycin: New agent PET imaging

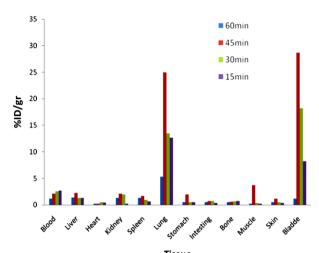


Figure 7. Biodistribution of ⁶⁸Ga-BLM in Bulb/c mice up to 60 min after injection.

DISCUSSION

In this research study, due the special properties of ⁶⁸Ga and BLM, the ⁶⁸Ga-BLM complex was prepared as a new PET imaging agent. The results of ⁶⁸Ge/⁶⁸Ga generator elution, the breakthrough of the generator and its possible chemical ionic impurities and the quality control of the eluted ⁶⁸GaCl₃ are very similar to the other literature ^(4,21).

The optimized conditions for radiolabeling of this radiolabeled complex were pH = 3.5-4; temperature = 90 °C; and reaction time = 15 min. The radiochemical purity of the complex was >96 % and the special activity of the final solution was > 17.4 GBq/mmol. The optimum reaction time for the complex found in this study agrees with Thakur method for labeling of 111In-BLM complex (14). Compared to the 177Lu-BLM (18) and 153Sm-BLM (19), the optimized pH for labeling of BLM with 177Lu and 153Sm (5.5 -7) has changed in labeling with 68Ga (3.5-4). Also, the labeling of BLM with 68Ga need to higher temperature (90 °C), however, the labeling time is shorter (15 min).

The biodistribution of 68 Ga-BLM in normal BALB/c mice showed different pattern compared with free 68 Ga cation. The water solubility of the cation cause its rapid removal from the blood circulation and the cation mainly extracted via urinary tract. The remained activity of 68 GaCl $_3$ in the body was accumulated

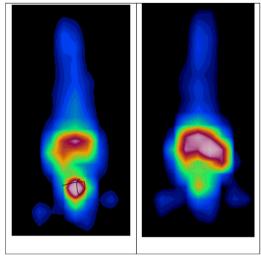


Figure 8. Images of ⁶⁸Ga-BLM 30 and 120 min post Injection.

in the stomach, spleen, liver and bone.

The biodistribution results of the radiolabeled compound clearly showed the main portion of the remained activity in the body was accumulated in lung and bladder. Also, the main extraction route of the compound is via the kidneys due the water solubility of the ⁶⁸Ga-BLM complex.

Since high resolution images for mice required an animal PET device and we did not have access to this instrument, the imaging was done by a human dual-head SPECT device, and therefore, the resulting images do not have suitable resolution, which is one of the limitations of this study. Other limitations include the lack of access to tumoral mice to investigate the biodistribution of radiolabeled compound in tumoral model. Given the anti-cancer properties of BLM on the one hand and the physical properties of gallium-68 on the other hand, the study of this radiolabeled compound in tumor model is suggested.

CONCLUSION

In this research study, ⁶⁸Ga-BLM was prepared in high radiochemical purity (>96 %) with high special activity (> 17.4 GBq/mmol) at a short time (15 min) using a portable ⁶⁸Ge/⁶⁸Ga generator. The biodistribution study of the radiolabeled complex showed completely

Int. J. Radiat. Res., Vol. 18 No. 2, April 2020

different pattern compared to the free gallium cation and lung and bladder were the accumulation sites for the remained activity. Generally, it seems ⁶⁸Ga-BLM can be considered as a high potential candidate for PET imaging; however, more biological studies in tumoral mammals are still needed.

ACKNOWLEDGMENTS

The authors gratefully acknowledge support from the Jaber Ibn Hayyan Laboratory Foundation of Nuclear Science and Technology Research Institute (NSTRI).

Conflicts of interest: Declared none.

REFERENCES

- 1. Velikyan I (2014) Prospective of ⁶⁸Ga-radiopharmaceutical development. *Theranostics*, **4:** 47-80.
- Firestone RB and Shirley VS (1998) Table of Isotopes, 2 Volume Set. In: Firestone RB, Shirley VS, editors. p. 3168. ISBN 0-471-33056-6. Wiley; 1998.
- Roesch F and Riss J (2010) The renaissance of the 68Ge/68Ga radionuclide generator initiates new developments in 68Ga radiopharmaceutical chemistry. Curr Top Med Chem, 10: 1633–1668.
- Sharifi M, Yousefnia H, Bahrami-Samani A, Jalilian AR, Zolghadri S, Alirezapour B, Geramifar P, Maus S, Beiki D (2017) Optimized production, quality control, biological evaluation and PET/CT imaging of ⁶⁸Ga-PSMA-617 in breast adenocarcinoma model. *Radiochim Acta*, 105: 399– 407.
- Rabie A, Yousefnia H, Zolghadri S, Shamsaei M, Jalilian AR (2018) Preparation, quality control and biodistribution study of ⁶⁸Ga-BPAMD: Optimized production with an inhouse ⁶⁸Ge-⁶⁸Ga generator. *Iran J Nucl Med*, 26: 82-86.
- Sattarzadeh E, Amini M.M, Kakaei S, Khanchi A (2018) ⁶⁸Ga -radiolabeled magnetic nanoparticles for PET–MRI imaging. J Radioanal Nucl Chem, 317, 3: 1333–1339.
- Pourmanouchehri Z, Jafarzadeh M, Kakaei S, Sattarzadeh E (2018) Magnetic Nanocarrier Containing ⁶⁸Ga–DTPA Com-

- plex for Targeted Delivery of Doxorubicin. *J Inorg Organ Poly Mat*, **28**(5): 1980–1990.
- Takita T, Muraoka Y, Nakatani T, Umezawa H (1978) Chemistry of bleomycin. XIX Revised structures of bleomycin and phleomycin. J Antibiot, 31: 801-804.
- Maeda K, Kosaka H, Yagashita K, Umezawa H (1957) Production and isolation of a new antibiotic: kanamycin. J Antibiot, 10: 181-188.
- 10. Umezawa H, Suhara Y, Takita T, Maeda K (1966) Purification of bleomycin. *J Antibiot*, **19**: 210-215.
- 11. Naganawa H, Muraoka Y, Takita T, Umezawa H (1977) Chemistry of Bleomycin. *J Antibiot*, *30*: 388-396.
- Burger RM, Peisach J, Horwitz SB (1981) Activated bleomycin. A transient complex of drug, iron and oxygen that degrades DNA. J Biol Chem, 256: 11636-11644.
- Brooks RC, Carnochan P, Sasabowski JK (1999) Metal complex of bleomycin: evaluation of (RH-105)-bleomycin for use in targeted radiotherapy. Nucl Med Biol, 26: 421-430.
- Jääskelä-Saari HA and Kairemo KJ (2005) Indium-111-Bleomycin complex in squamous cell cancer xenograft tumors of nude mice. Cancer Biother Radio, 20: 426-435.
- Woolfenden JM, Alberts DS, Hall JN, Patton DD (1979) Cobalt-57 bleomycin for imaging head and neck tumors. Cancer, 43: 1652-1657.
- 16 Ugur O, Caner B, Balbay MD, Remzi D (1993) Bleomycin lung toxicity detected by technetium-99m diethylene triamine penta-acetic acid aerosol scintigraphy. Eur J Nucl Med, 20: 114-118.
- Padbury G and Ferric J (1988) Bleomycin catalyzed reduction of 10-hydroperoxy-8, 12-octadecadienoic acid: evidence for hemolytic oxygen-oxygen bond scission. *Biochemistry*, 27: 7846–7852.
- Yousefnia H, Jalilian AR, Zolghadri S, Bahrami-Samani A, Shirvani-Arani S, Ghannadi-Maragheh M (2010) Preparation and quality control of lutetium-177 bleomycin as a possible therapeutic agent. Nukleonika, 55: 285–291.
- Bahrami-Samani A, Ghanadi-Maragheh M, Jalilian AR, Yousefnia H, Moradkhani S, Bolourinovin F (2010) Development of ¹⁵³Sm-bleomycin as a possible therapeutic complex. *Nucl Sci Tech*, 21: 16–19.
- Zolghadri S, Jalilian AR, Yousefnia H, Bahrami-Samani A, Shirvani-Arani S, Ghannadi-Maragheh M (2010) Development of 166Ho bleomycin as a possible therapeutic complex. J Radioanal Nucl Chem, 285: 461–467.
- Mirzaei A, Jalilian A, Aghanejad A, Mazidi M, Yousefnia H, Beiki D (2015) Preparation and evaluation of ⁶⁸Ga-ECC as a PET renal imaging agent. *Nucl Med Mol Imaging*, 49: 208-216.
- 22. IAEA (2004) IAEA-TECDOC-1401: Quantifying Uncertainty in Nuclear Analytical Measurements. IAEA, Vienna, Austria