Iodine-125 radiation inhibits epithelial-mesenchymal transition in lung cancer cells by blocking TGF-β1/Smad3/Snai1 signaling

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ABSTRACT

Background: Iodine-125 (125I) brachytherapy is an effective strategy for treating human tumors. The current study aimed to discover the mechanisms underlying the therapeutic action of ¹²⁵I radiation in lung cancer, with a focus on its impact on the epithelial-mesenchymal transition (EMT). Materials and Methods: A549 cells, a human lung adenocarcinoma cell line, were treated with transforming growth factor $\beta 1$ (TGF- $\beta 1$) and/or ¹²⁵I (control, TGF- $\beta 1$, ¹²⁵I, and TGF- β 1 + 125 I groups) to evaluate the effects of 125 I on TGF- β 1-induced EMT. After treatment, the viability of A549 cells was detected using 3-(4,5dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. The expression of E-cadherin, N-cadherin, Vimentin, Smad3, and Snai1 markers and pathway molecules was measured. Results: 125 radiation inhibited the viability of A549 cells, both with and without TGF-β1 treatment. TGF-β1 intervention promoted the EMT of A549 cells, as demonstrated by the morphological transition from a polygonal shape to a spindle shape, reduced E-cadherin levels, and elevated Vimentin and N-cadherin expression. Notably, TGF-β1-activated EMT was significantly weakened by ¹²⁵I radiation. Moreover, ¹²⁵I radiation reversed TGF-β1-induced upregulation of Smad3 and Snai1 in A549 cells. *Conclusion:* 125 I radiation suppresses the EMT by blocking TGF- β 1/ Smad3/Snai1 signaling, contributing to the treatment of lung cancer.

INTRODUCTION

Lung cancer is a common and fatal malignancy worldwide (1,2). According to global cancer statistics, in 2022, there were 2,480,301 newly diagnosed cases and 1,817,172 deaths, cases in 2022, making lung cancer the leading cause of both cancer incidence and mortality worldwide (3). Smoking is the main cause of 80-90% of lung cancer cases, with other factors including radon or fume exposure, toxic occupational environment, and microbial infection, among others (4, 5). Similar to other types of cancer, surgery, chemotherapy, and radiotherapy are traditional therapeutic strategies for lung cancer in clinical practice (6). The development of immunotherapy and molecular targeted therapy has great advantages for improving the outcomes of patients with lung cancer (7). However, the outcome of lung cancer remains poor, particularly because of late diagnosis, metastasis, and drug resistance (8,9). Therefore, more effective therapeutic strategies are required for lung

Brachytherapy is a type of radiotherapy that puts radioactive sources within or close to the tumor parenchyma, allowing more concentrated radiotherapy while sparing surrounding normal tissues (10). Evidence has shown that brachytherapy is effective and safe for tumors located at various sites, such as the lungs, liver, colon, pancreas, prostate, breast, and endometrium (11, 12). In brachytherapy, radioactive sources contain Cobalt-60, iridium-192, ruthenium-106, palladium-103, and astatine-211 (13, 14), among which iodine-125 (125I) is the most common one (15-17). 125I brachytherapy is widely used to treat diverse types of tumors (16, 18-20), including lung cancer (21). A biocentric analysis showed that 125I brachytherapy was more effective in elevating the survival rate within six months without progression (76.3% vs. 51.5%, P=0.002) than external beam radiotherapy in patients with non-small cell lung cancer (NSCLC) accompanied by brain metastasis (22). ¹²⁵I brachytherapy improves clinical outcomes and reduces the possibility of myelosuppression in patients when compared with chemotherapy (23). Another meta-analysis involving eight studies found that ¹²⁵I brachytherapy improved the efficacy of transarterial chemical infusion without inducing severe adverse events in patients with advanced lung cancer ⁽²⁴⁾. However, the therapeutic mechanisms of ¹²⁵I brachytherapy in lung cancer have not been fully elucidated.

Epithelial-mesenchymal transition (EMT), a process involving cell morphological transformation, plays a crucial role not only in embryogenesis and wound healing but also in tumour progression (25, 26). In terms of molecular mechanisms, transcription factors, including SNAI, ZEB, and TWIST, are master regulators of EMT (27). The TGF-β pathway is a predominant signalling pathway involved in the regulation of EMT. As a major driver, TGF-β can induce EMT via activating Smad/non-Smad and cross -talking with many other signals (28). Recent studies have revealed that many agents exhibit therapeutic potential in lung cancer by blocking TGF-β-mediated EMT. For example, salinomycin inhibits cell migration and invasion by suppressing TGF-β1-activatied EMT in lung cancer. Cucurbitacin B inhibits TGF-β1activated EMT in NSCLC, contributing to remission of tumour metastasis in vivo (29). A TGF-β1 inhibitor, Compound 67 (an analogue of chalcones), suppresses cell migration, invasion, and EMT in lung cancer (30).

The above demonstrates that EMT is closely linked to metastasis and recurrence of lung adenocarcinoma and that 125I radiation exerts superior effects in patients with advanced lung adenocarcinoma. However, there are few studies on the association between radioactive 125I particles and EMT, as well as the underlying mechanism in the context of lung adenocarcinoma. Hence, we aimed to explore the mechanism of action of 125I radiation in lung cancer, primarily concerning EMT. Specifically, TGF-β1 was used to induce EMT in human lung adenocarcinoma cells. The effects of 125I radiation on EMT and TGF-β1/Smad3/Snai1 signaling were mainly explored at the cellular level. This study aims to reveal the mechanism of action of 125I radiation on EMT related to the TGF-β1/Smad3/Snai1 signaling, laying the foundation for lung cancer therapy.

MATERIALS AND METHODS

Cell culturing and treatments

A549 cells (a human lung adenocarcinoma cell line; Procell, Wuhan, China) were maintained in RPMI -1640 medium (Meilun, Xingtai, China) containing 10% foetal bovine serum (FBS; Gibco, Grand Island, NY, USA) at 37 °C with 5% CO₂.

 125 I seeds were obtained from Beike Biotechnology (Beijing, China) and stored on lead clips. The parameters of 125 I seeds are as follows: diameter = 0.8 mm, length = 4.5 mm, effective radiation radius = 17 mm, half-life = 59.43 d, X-ray energy = 27.4-31.5 keV, γ radiation energy = 35.5

keV, and radiation activity = 0.8 mCi. For treatment, A549 cells were categorised into four groups. In the TGF- β 1 group, cells received 48 h of treatment with 5 ng/mL TGF- β 1, as previously described ⁽³¹⁾. In the ¹²⁵I group, cells were irradiated with 10 ¹²⁵I seeds at an interval of 1 cm. In the TGF- β 1 + ¹²⁵I group, cells received both TGF- β 1 treatment and ¹²⁵I irradiation simultaneously. Untreated cells were used as the control group. After treatment, the morphological transformation of A549 cells was monitored under a microscope (MOTIC, Chengdu, China).

Throughout the experiment, the personnel were equipped with lead-protective clothing. Cells in each group were cultured in a separate incubator and separated with 0.5 mm lead plates. After the experiments, all ¹²⁵I seeds were recycled into a lead tank and disposed of according to the guidelines for radioactive waste.

MTT analysis

The viability of A549 cells in different groups was determined using an MTT assay kit (Sparkjade, Qingdao, China) according to the manufacturer's instructions. Simply, we seeded A549 cells (2×10⁴ cells/well) in 96-well plates and cultured for 24 h. Afterwards, cells were stimulated with TGF- β 1 and/or 125 I for another 24 or 48 h (the 48th and 72th hour). After 4 h of incubation with 10 μ L MTT working solution, cells were continuously treated with 100 μ L dimethyl sulfoxide for 2 h. The optical density was measured at 570 nm and was eventually read by a microplate reader (Hiwell Diatek, Wuxi, China).

qRT-PCR

Cells were lysed in TRIzol reagent (Vicbio Biotechnology Co., Ltd., Beijing, China) on ice to separate RNA samples, and the isolated RNAs were immediately reverse-transcribed into cDNAs using a cDNA synthesis kit (Servicebio, Wuhan, China). Using a SYBR Green qPCR Master Mix (Servicebio, Wuhan, China), qRT-PCR was run on a PCRAmplifier (#CFX96 Touch Deep Well, Bio-Rad, CA, USA). Utilising the 2-\(^{\text{DACt}}\) method (internal control: GAPDH), we calculated the relative expression of specific mRNAs. The primer sequences are listed in table 1.

Table 1. The used primers in this study.

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Name	Gene symbol	Gene ID	
E- cadherin	CDH1	999	Forward primer: TGGACCGAGA- GAGTTTCCCT
			Reverse primer: TTAGCCTCGTTCTCAGGCAC
Vimentin	VIM	7431	Forward primer: GGAC- CAGCTAACCAACGACA
			Reverse primer: AAGGTCAA- GACGTGCCAGAG
N- cadherin	CDH2	1000	Forward primer: GAGGCTTCTGGTGAAATCGC
			Reverse primer: AATCTG- CAGGCTCACTGCTC
SMAD3	SMAD3	4088	Forward primer: TCCATGACTGTGGATGGCTTC
			Reverse primer: TTCAGGTT- GCATCCGATGTG
SNAI1	SNAI1	6615	Forward primer: TAGCGAG- TGGTTCTTCTGCG
			Reverse primer: AGGGCTGCTG- GAAGGTAAAC

Western blot

Cells were lysed in RIPA lysis buffer (Servicebio, Wuhan, China) on ice to isolate protein samples. Equivalent amounts of protein from different groups were used for detection. First, after separation using 10% SDS-PAGE, we transferred the proteins onto PVDF membranes (Millipore Billerica, MA, USA). Subsequently, the membrane was blocked with 5% BSA for 30 min at 37 °C, followed by co-incubation with the following primary antibodies (anti-β-actin, Wuhan, Proteintech, China: anti-E-cadherin, -Vimentin, -N-cadherin, -Smad3, and -Snai1, Abcam, Cambridge, UK) for 12 h at 4 °C. Afterwards, coincubation with secondary antibody was carried out for 1 h (HRP-labelled IgG, Servicebio, Wuhan, China) at 25 °C. Finally, the membranes were visualised using an efficient chemiluminescence kit (Servicebio, Wuhan, China) and quantified using a Gel Imaging System (Tanon, China).

Statistical analysis

Quantitative data were displayed as mean ± standard deviation and were analyzed using GraphPad Prism v7.0 (San Diego, CA, USA). One-way ANOVA and Tukey's test were used to compare data across the four groups. Statistical significance was set at P<0.05.

RESULTS

Radiation from ¹²⁵I inhibits the viability of A549 cells

The viability of the A549 cells in response to 125 I radiation was analyzed. At both 48 and 72 h post-treatment, a significantly higher viability was observed in TGF- β 1-stimulated A549 cells in comparison with non-stimulated A549 cells (P<0.01). 125 I radiation decreased the viability of A549 cells, regardless of TGF- β 1 stimulation, with significant levels (P<0.01) (figure 1).

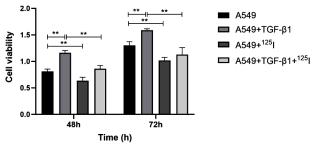


Figure 1. The viability of A549 cells in different groups was measured by MTT assay. A549 cells were treated with TGF-β1 and/or 125I. **P < 0.01.

Radiation from ¹²⁵I inhibits the EMT of A549 cells

Three EMT markers were measured to determine the effects of ¹²⁵I radiation on EMT. As displayed in

figure 2a, TGF- β 1 induced a down-regulation of E-cadherin mRNA expression in A549 cells (P<0.05). On the contrary, mRNA levels of Vimentin and N-cadherin were enhanced in TGF- β 1-stimulated A549 cells in comparison with non-stimulated A549 cells (P<0.05) (figuree 2b and c). Radiation from ¹²⁵I significantly elevated E-cadherin levels while decreasing Vimentin and N-cadherin levels in A549 cells (P<0.01). Additionally, it reversed the effects of TGF- β 1 on the regulating of these EMT markers (P<0.05) (figure 2a-c). In accordance with the above observations on mRNA levels, western blotting revealed the same changes in the protein levels of E-cadherin, N-cadherin, and Vimentin in the different groups (P<0.05) (figure 2d-g).

Radiation from ¹²⁵I blocks TGF-β1/Smad3/Snai1 signaling in A549 cells

The mechanism of action of 125 I radiation on Smad3 and Snai1 cells was further evaluated. As a result, the mRNA and protein levels of Smad3 and Snai1 were elevated by TGF- β 1 (P < 0.05) but were decreased by 125 I radiation in A549 cells (P<0.01). Notably, the upregulation of Smad3 and Snai1 induced by TGF- β 1 was significantly weakened by 125 I radiation (P<0.05) (figure 3a-e).

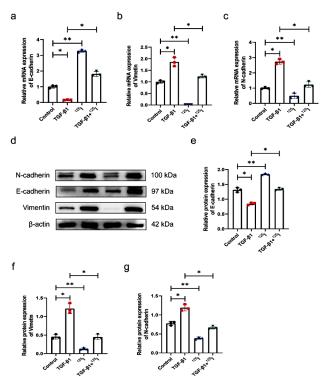
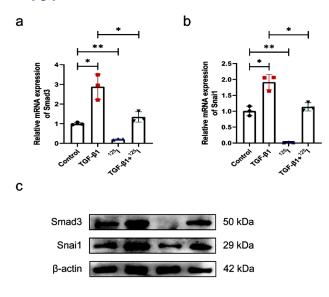


Figure 2. The expression of three EMT markers in A549 cells of different groups. a, The mRNA expression of E-cadherin; b, The mRNA expression of Vimentin; c, The mRNA expression of N-cadherin; d-g, The protein expression of E-cadherin, Vimentin, and N-cadherin. *P < 0.05, **P < 0.01.



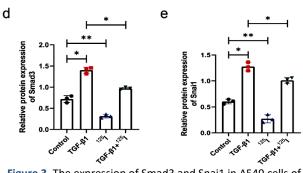


Figure 3. The expression of Smad3 and Snai1 in A549 cells of different groups. **a,** The mRNA expression of Smad3; **b,** The mRNA expression of Snai1; **c-e,** The protein expression of Smad3 and Snai1; *P < 0.05, **P < 0.01.

DISCUSSION

Lung cancer is a common pulmonary tumor associated with high mortality rates worldwide (32). As a minimally invasive radiotherapy, brachytherapy can be used as an alternative therapy for lung cancer at an early stage and as a principal or auxiliary therapy for lung cancer at a locally advanced stage or with metastasis (33). However, knowledge of the mechanisms of action of brachytherapy involving EMT in lung cancer remains limited.

EMT is an important biological process that drives the epithelial-to-mesenchymal phenotype. In addition to embryonic development, wound healing, and organ fibrosis, EMT also plays a prominent role in metastasis and resistance (34, 35). On the one hand, EMT endows tumor cells with the abilities of stemness and invasiveness, contributing to distant metastasis and recurrence (36). On the other hand, EMT also enhances the drug efflux pump and antiapoptotic capacity of tumor cells, contributing to drug resistance (34). Therefore, targeting the EMT is a key focus in cancer treatment. Evidence has determined that A variety of molecules, including

EFEMP2 (37), ALOX12 (38), PTPL1 (39), HRH3 (40), and LINC00891 (41), act as potential therapeutic targets for lung cancer by inhibiting EMT, such as EFEMP2 (37), ALOX12 (38), PTPL1 (39), HRH3 (40), and LINCO0891 (41), among others. One review summarized that natural product-derived compounds with the ability to inhibit EMT are beneficial for the treatment of lung cancer (42). E-cadherin, Vimentin and N-cadherin are wellknown biomarkers of EMT; E-cadherin is a marker for epithelial cells, whereas the other two are markers for mesenchymal cells (43). As a typical epithelial cell marker of EMT, downregulation of E-cadherin can induce EMT in tumor cells and promote their metastasis (44). E-cadherin expression is enhanced in human liver cancer cells after radioactive 125I treatment (45). Metastasis of cancer cells is highly correlated with vimentin expression in NSCLC (46). Abnormal expression of N-cadherin enhances tumor cell invasion and migration, and N-cadherin-induced MMP-9 is regarded as a core step in tumor invasion and metastasis (47). In TGF-β1-treated cells, 125I radiation increased E-cadherin levels while reducing Vimentin and N-cadherin levels, indicating inhibition of EMT after 125I therapy. Similar results were observed in a previous study (48).

TGF- β is a pivotal cytokine that is implicated in the of modulation embryonic development, organogenesis, wound healing, immunomodulation, fibrosis, and carcinogenesis, among others (49). Notably, TGF-β is well-known as an inducer of EMT, and its activation is positively associated with metastasis and chemotherapy resistance in cancer (50). The underlying mechanisms of TGF-β in EMT are complex, and the TGF-β1/Smad3 signal axis is a pivotal one. In cells, the receptor complex of TGF-β1 can lead to the activation of Smad2/3 by inducing Cterminal phosphorylation. The trimers formed by Smad2, 3 and 4 then bind to DNA-binding transcription factors and synergistically regulate the target genes involved in EMT (28). Until now, emerging evidence has determined that TGF-β1/Smad3 signaling is the target of agents with therapeutic potential in cancer. For example, baicalin inhibits EMT-related metastasis in breast cancer by inhibiting TGF-β1/Smad3 signaling (51). Nobiletin inhibits the EMT of NSCLC cells by blocking TGF-β1/Smad3 signaling (52). OPB suppresses EMT in TGF-β1-induced lung cancer cells by regulating Smad3 (53). Snai1 is also a pivotal modulator of EMT and can be activated by Smad3 (54), which induces EMT by directly inhibiting E-cadherin transcription Overexpression of Snail1 downregulates E-cadherin and Plakoglobin, and upregulates Vimentin and Fibronectin to activate EMT (28). Therefore, the TGFβ1/Smad3/Snai1 signaling pathway is an important mechanism by which 125I radiation inhibits EMT in lung cancer.

Our study demonstrates that the therapeutic action of ¹²⁵I radiation in lung cancer is partly

achieved by inhibiting EMT. However, several limitations of this study should be considered. EMT is closely associated with tumor metastasis, and we mainly focused on the effects of ¹²⁵I radiation on EMT marker expression, while ignoring its effects on tumor cell migration and invasion. In addition, this was a preliminary study that included only a limited number of in vitro experiments. Further in vivo experiments are required to verify the findings obtained from this study.

CONCLUSION

In conclusion, TGF- β 1 induces the EMT in lung cancer cells, an effect that is reversed by ¹²⁵I radiation. This radiation may inhibit EMT by blocking the TGF- β 1/Smad3/Snai1 signaling pathway.

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study.

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Author contribution: Conception and design of the research: F.S., H.L. and D.X.; Acquisition of data: J.C., M.H. and G.Z.; Analysis and interpretation of data: G.Z.; Statistical analysis: H.L. and D.X.

REFERENCES

- Thai AA, Solomon BJ, Sequist LV, Gainor JF, Heist RS (2021) Lung cancer. Lancet (London, England), 398(10299): 535-54.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. (2021) Global Cancer Statistics 2020: GLOBOCAN Estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin, 71(3): 209-49.
- Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. (2024) Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: a Cancer Journal for Clinicians, 74(3): 229-263.
- Schabath MB and Cote ML (2019) Cancer Progress and Priorities: Lung Cancer. Cancer Epidemiol Biomarkers Prev, 28(10): 1563-79.
- Corrales L, Rosell R, Cardona AF, Martin C, Zatarain-Barron ZL, Arrieta O (2020) Lung cancer in never smokers: The role of different risk factors other than tobacco smoking. Crit Rev Oncol Hematol, 148: 102895.
- Xue T, Zhao X, Zhao K, Lu Y, Yao J, Ji X (2022) Immunotherapy for lung cancer: Focusing on chimeric antigen receptor (CAR)-T cell therapy. Curr Probl Cancer, 46(1): 100791.
- 7. Alexander M, Kim SY, Cheng H (2020) Update 2020: Management of non-small cell lung cancer. *Lung*, **198**(6): 897-907.
- Denisenko TV, Budkevich IN, Zhivotovsky B (2018) Cell death-based treatment of lung adenocarcinoma. Cell Death Dis, 9(2): 117.
- Herbst RS, Morgensztern D, Boshoff C (2018) The biology and management of non-small cell lung cancer. Nature, 553(7689): 446-54.
- 10. Otter SJ, Stewart AJ, Devlin PM (2019) Modern Brachytherapy. Hematol Oncol Clin North Am, 33(6): 1011-25.
- 11. Deng X, Wu H, Gao F, Su Y, Li Q, Liu S, et al. (2017) Brachytherapy in the treatment of breast cancer. *Int J Clin Oncol*, 22(4): 641-50.

- 12. Takeshita A, Shimamoto H, Uchimoto Y, Tsujimoto T, Miyamoto T, Kreiborg S, et al. (2023) High-dose-rate mold brachytherapy for mandibular gingival squamous cell carcinoma in outpatient setting-lnitial case report. *International Journal of Radiation Research*, 21 (4): 849-53.
- 13.Li Y, Pang F, Cai H, Li L (2022) Intratumoral injection of radioactive Astatine (211At) microspheres for the treatment of tumors. *International Journal of Radiation Research*, 20(4): 793-8.
- 14. Muñoz Arango E, Pickler A, Mantuano A, Salata C, de Almeida CE (2020) Feasibility study of the Fricke chemical dosimeter as an independent dosimetric system for the small animal radiation research platform (SARRP). *Physica Medica*, 71: 168-75.
- 15. Kaliki S and Shields CL (2017) Uveal melanoma: relatively rare but deadly cancer. Eye (Lond), 31(2): 241-57.
- 16. Gao H, Wang Y, Du C, Li X, Liu K, Xue H, et al. (2022) Study on the factors affecting the dose error of using I-125 seeds in the treatment of prostate cancer using the Monte Carlo method. *International Journal of Radiation Research*, 20(4): 857-64.
- 17. Ghahramani MR, Asgharizadeh F, Assadi MR, Ahmadi SJ, Moradi K (2012) Effects of different carriers for adsorption of I-125 on brachytherapy sources. *International Journal of Radiation Research*, 10(2): 105-7.
- Yu H, Zhang H, Gao Z, Liu X, Zhang L, Di X, et al. (2022) (125)I Seed brachytherapy for refractory loco-regional recurrence of nonanaplastic thyroid cancer. Frontiers in Oncology, 12: 773708.
- Huang S, Cao Y, Wang R, Liu H, Wang T, Yang S (2023) Feasibility of 125I brachytherapy combined with arterial infusion chemotherapy in patients with advanced pancreatic cancer. *Medicine*, 102(44): e35033.
- 20. Yao X, Lu S, Feng C, Suo R, Li H, Zhang Y, et al. (2022) Tumor oxygenation nanoliposome synergistic hypoxia-inducible-factor-1 inhibitor enhanced Iodine-125 seed brachytherapy for esophageal cancer. *Biomaterials*, 289: 121801.
- 21. Wang Y, Li F, Hu Y, Sun Y, Tian C, Cao Y, et al. (2023) Clinical outcomes of intra-arterial chemotherapy combined with iodine-125 seed brachytherapy in the treatment of malignant superior vena cava syndrome caused by small cell lung cancer. Cancer Radiotherapie, 27(4): 312-8.
- Yang L, Wang C, Zhang W, Liu S, Xuan T, Jiang H, et al. (2022) lodine
 -125 brachytherapy treatment for newly diagnosed brain metastasis in non-small cell lung cancer: A biocentric analysis. *Front Oncol*, 12: 1005876.
- Zhang W, Li J, Li R, Zhang Y, Han M, Ma W (2018) Efficacy and safety of iodine-125 radioactive seeds brachytherapy for advanced nonsmall cell lung cancer-A meta-analysis. *Brachytherapy*, 17(2): 439-490
- 24. Hong J, Shi YB, Fu YF, Yang LL (2022) Iodine-125 seeds insertion with trans-arterial chemical infusion for advanced lung cancer: a meta-analysis. J Contemp Brachytherapy, 14(4): 403-10.
- Lamouille S, Xu J, Derynck R (2014) Molecular mechanisms of epithelial-mesenchymal transition. Nat Rev Mol Cell Biol, 15(3): 178-96.
- 26. Dongre A and Weinberg RA (2019) New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer. Nat Rev Mol Cell Biol, 20(2): 69-84.
- Debnath P, Huirem RS, Dutta P, Palchaudhuri S (2022) Epithelial-mesenchymal transition and its transcription factors. *Biosci Rep*, 42 (1): BSR20211754.
- Xu J, Lamouille S, Derynck R (2009) TGF-beta-induced epithelial to mesenchymal transition. Cell Res, 19(2): 156-72.
- 29. Yuan R, Fan Q, Liang X, Han S, He J, Wang QQ, et al. (2022) Cucurbitacin B inhibits TGF-beta1-induced epithelial-mesenchymal transition (EMT) in NSCLC through regulating ROS and PI3K/Akt/mTOR pathways. *Chin Med*, *17*(1): 24.
- 30. Jeong JH, Jang HJ, Kwak S, Sung GJ, Park SH, Song JH, et al. (2019) Novel TGF-beta1 inhibitor antagonizes TGF-beta1-induced epithelial-mesenchymal transition in human A549 lung cancer cells. J Cell Biochem. 120(1): 977-87.
- Kim J and Hwan Kim S (2013) CK2 inhibitor CX-4945 blocks TGF-β1induced epithelial-to-mesenchymal transition in A549 human lung adenocarcinoma cells. PloS One, 8(9): e74342.
- 32. Amiri A, Pourhanifeh MH, Mirzaei HR, Nahand JS, Moghoofei M, Sahebnasagh R, et al. (2021) Exosomes and lung cancer: roles in pathophysiology, diagnosis and therapeutic applications. *Curr Med Chem*, 28(2): 308-28.
- 33. Qiu B, Jiang P, Ji Z, Huo X, Sun H, Wang J (2021) Brachytherapy for lung cancer. *Brachytherapy*, 20(2): 454-66.

- 34. Du B and Shim JS (2016) Targeting Epithelial-mesenchymal transition (EMT) to overcome drug resistance in cancer. *Molecules, 21*
- 35. Roche J (2018) The epithelial-to-mesenchymal transition in cancer. *Cancers (Basel)*, 10(2): 52.
- Zhang Y and Weinberg RA (2018) Epithelial-to-mesenchymal transition in cancer: complexity and opportunities. Front Med, 12(4): 361-73.
- 37. Song L, Li XX, Liu XY, Wang Z, Yu Y, Shi M, et al. (2020) EFEMP2 Suppresses the invasion of lung cancer cells by inhibiting epithelial-mesenchymal transition (EMT) and down-regulating MMPs. *Onco Targets Ther*, **13**: 1375-96.
- Chen J, Tong W, Liao M, Chen D (2020) Inhibition of arachidonate lipoxygenase12 targets lung cancer through inhibiting EMT and suppressing RhoA and NF-kappaB activity. *Biochem Biophys Res* Commun, 524(4): 803-9.
- 39.Zhu N, Zhang XJ, Zou H, Zhang YY, Xia JW, Zhang P, et al. (2021) PTPL1 suppresses lung cancer cell migration via inhibiting TGFbeta1-induced activation of p38 MAPK and Smad 2/3 pathways and EMT. Acta Pharmacol Sin, 42(8): 1280-7.
- 40. Zhao YY, Jia J, Zhang JJ, Xun YP, Xie SJ, Liang JF, et al. (2021) Inhibition of histamine receptor H3 suppresses the growth and metastasis of human non-small cell lung cancer cells via inhibiting PI3K/Akt/mTOR and MEK/ERK signaling pathways and blocking EMT. Acta Pharmacol Sin, 42(8): 1288-97.
- 41.Zhang Y, Song D, Peng Z, Wang R, Li K, Ren H, et al. (2022) LINC00891 regulated by miR-128-3p/GATA2 axis impedes lung cancer cell proliferation, invasion and EMT by inhibiting RhoA pathway. Acta Biochim Biophys Sin (Shanghai), 54(3): 378-87.
- Chanvorachote P, Petsri K, Thongsom S (2022) Epithelial to Mesenchymal Transition in Lung Cancer: Potential EMT-targeting natural product-derived compounds. *Anticancer Res.* 42(9): 4237-46.
- Odero-Marah V, Hawsawi O, Henderson V, Sweeney J (2018) Epithelial-mesenchymal transition (EMT) and prostate cancer. Adv Exp Med Biol, 1095: 101-10.
- 44. Li Q, Zhou ZW, Duan W, Qian CY, Wang SN, Deng MS, et al. (2021) Inhibiting the redox function of APE1 suppresses cervical cancer metastasis via disengagement of ZEB1 from E-cadherin in EMT. J Exp Clin Can Res, 40(1): 220.

- 45. Yang C, Xiao Y, Du Y, Xiong J, Deng L, Liang Q, et al. (2022) Iodine-125 seeds inhibit carcinogenesis of hepatocellular carcinoma cells by suppressing epithelial-mesenchymal transition via TGF-β1/Smad signaling. *Disease Markers*, 2022: 9230647.
- 46. Berr AL, Wiese K, Dos Santos G, Koch CM, Anekalla KR, Kidd M, et al. (2023) Vimentin is required for tumor progression and metastasis in a mouse model of non-small cell lung cancer. Oncogene, 42 (25): 2074-87.
- 47. Cao ZQ, Wang Z, Leng P (2019) Aberrant N-cadherin expression in cancer. *Biomedicine & pharmacotherapy*, **118**: 109320.
- He Y, Li L, Liu J, Zhang X (2018) Iodine-125 seed brachytherapy inhibits non-small cell lung cancer by suppressing epithelial-mesenchymal transition. *Brachytherapy*, 17(4): 696-701.
- Peng D, Fu M, Wang M, Wei Y, Wei X (2022) Targeting TGF-beta signal transduction for fibrosis and cancer therapy. *Mol Cancer*, 21 (1): 104.
- Hao Y, Baker D, Ten Dijke P (2019) TGF-beta-Mediated Epithelial-Mesenchymal Transition and Cancer Metastasis. Int J Mol Sci, 20 (11): 2767.
- 51. Liu DK, Dong HF, Liu RF, Xiao XL (2020) Baicalin inhibits the TGF-beta1/p-Smad3 pathway to suppress epithelial-mesenchymal transition-induced metastasis in breast cancer. *Oncotarget*, 11(29): 2863-72.
- 52. Da C, Liu Y, Zhan Y, Liu K, Wang R (2016) Nobiletin inhibits epithelial -mesenchymal transition of human non-small cell lung cancer cells by antagonizing the TGF-beta1/Smad3 signaling pathway. *Oncol Rep*, 35(5): 2767-74.
- 53. Liu X, Zhang Y, Zhou GJ, Hou Y, Kong Q, Lu JJ, et al. (2020) Natural alkaloid 8-oxo-epiberberine inhibited TGF-beta1-triggred epithelialmesenchymal transition by interfering Smad3. *Toxicol Appl Phar*macol, 404: 115179.
- 54. Cho HJ, Baek KE, Saika S, Jeong MJ, Yoo J (2007) Snail is required for transforming growth factor-beta-induced epithelial-mesenchymal transition by activating PI3 kinase/Akt signal pathway. *Biochem Biophys Res Commun*, 353(2): 337-43.
- 55. Serrano-Gomez SJ, Maziveyi M, Alahari SK (2016) Regulation of epithelial-mesenchymal transition through epigenetic and post-translational modifications. *Molecular Cancer*, 15: 18.